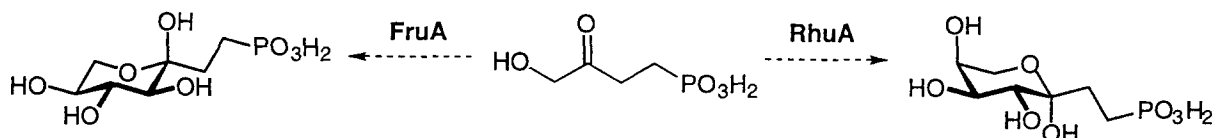


Carbohydr. Res. **1997**, 305, 313

Practical synthesis of 4-hydroxy-3-oxobutylphosphonic acid and its evaluation as a bio-isosteric substrate of DHAP aldolases

Hans-Lothar Arth, Wolf-Dieter Fessner

Department of Organic Chemistry, RWTH Aachen, D-52056 Aachen, Germany



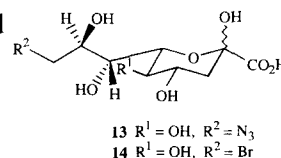
Carbohydr. Res. **1997**, 305, 323

The chemoenzymatic synthesis of 9-substituted 3,9-di-deoxy-D-glyxero-D-galacto-2-nonulosonic acids

David C.M. Kong, Mark von Itzstein

Department of Medicinal Chemistry, Victorian College of Pharmacy, Monash University (Parkville Campus), 381 Royal Parade, Parkville, Victoria 3052, Australia

The preparation of some novel C-9 modified Kdn analogues has been achieved from the corresponding 6-modified mannoses using Neu5Ac aldolase.

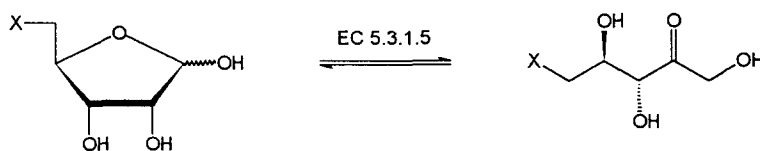


Carbohydr. Res. **1997**, 305, 331

Glucose isomerase catalysed isomerisation reactions of (2R,3R)-configured alsofuranoses into the corresponding open-chain 2-ketoses

Michael Ebner, Arnold E. Stütz

Institut für Organische Chemie der Technischen Universität Graz, Stremayrgasse 16, A-8010 Graz, Austria



Carbohydr. Res. **1997**, 305, 337

Enzymatic synthesis of L-tagatose from galactitol with galactitol dehydrogenase from *Rhodobacter sphaeroides* D

Alexander Huwig, Susanne Emmel, Gregor Jäkel, Friedrich Giffhorn

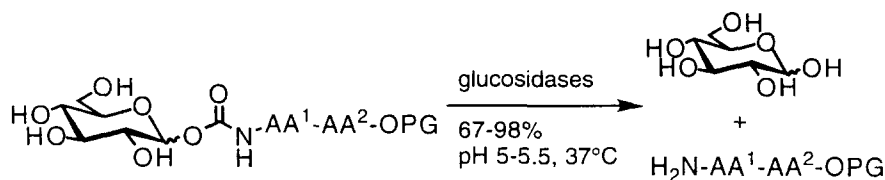
Inst. Applied Microbiology, Saarbrücken, Germany

Bioconversion of galactitol **1** to L-tagatose **2** by galactitol dehydrogenase from *Rhodobacter sphaeroides* D. Operational conditions are described in *Experimental*.

The tetrabenzylglucosyloxycarbonyl(BGloc)-group a carbohydrate-derived enzyme-labile urethane protecting group

Thomas Kappes, Herbert Waldmann

Institut für Organische Chemie, Universität Karlsruhe, Richard-Willsätter-Allee 2, D-76128 Karlsruhe, Germany



Diastereoselective resolution of 6-substituted glycosides via enzymatic hydrolysis

Urszula Grabowska ^c, David A. MacManus ^b, Keith Biggadike ^a, Mike I. Bird ^a, Stephen Davies ^b, Timothy Gallagher ^c, Lee D. Hall ^d, Evgeny N. Vulfson ^b

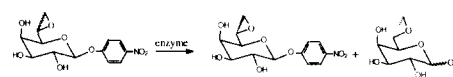
^a GlaxoWellcome, Medicines Research Centre, Stevenage, SG1 2NY, UK

^b Food Macromolecular Sciences Department, Institute of Food Research, Earley Gate, Whiteknights Road, Reading, RG6 6BZ, UK

^c School of Chemistry, University of Bristol, Bristol, BS8 1TS, UK

^d X-Ray Crystallographic Unit, School of Chemistry, University of Bristol, Bristol, BS8 1TS, UK

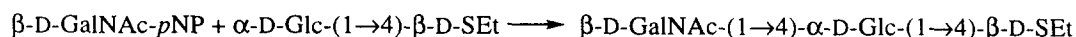
The enzymatic resolution of diastereomeric β -D-galactosides incorporating a variety of stereogenic centres at the 6-position is described.



Trisaccharide synthesis by glycosyl transfer from *p*-nitrophenyl β -D-N-acetylgalactosaminide on to disaccharide acceptors catalysed by the β -N-acetylhexosaminidase from *Aspergillus oryzae*

Suddham Singh, Michaela Scigelova, Peter Critchley, David H.G. Crout

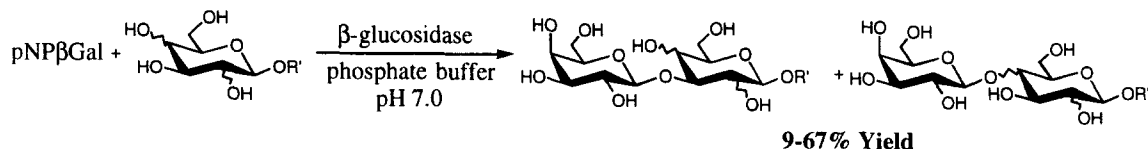
Department of Chemistry, University of Warwick, Coventry CV4 7AL, UK



Enzymatic synthesis of disaccharides using *Agrobacterium* sp. β -glucosidase

Heiko Prade, Lloyd F. Mackenzie, Stephen G. Withers

Protein Engineering Network of Centres of Excellence of Canada and Department of Chemistry, University of British Columbia, 2036 Main Mall, Vancouver, British Columbia, Canada, V6T 1Z1



Regioselectivity of the enzymatic transgalactosidation of D- and L-xylose catalysed by β -galactosidases

Esther Montero ¹, Jose Alonso ¹, Francisco J. Cañada ¹, Alfonso Fernández--Mayoralas ¹, Manuel Martín-Lomas ²

¹ Instituto de Química Orgánica, CSIC, Juan de la Cierva 3, 28006 Madrid, Spain

² Instituto de Investigaciones Químicas, CSIC, Isla de la Cartuja, Sevilla, Spain

The synthesis of β -D-galactopyranosyl-xyloses using different β -galactosidases is described.

β -D-Gal-ONP + Xylose $\xrightarrow{\beta\text{-Galactosidase}}$

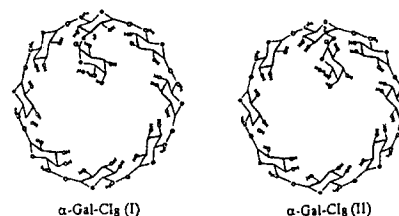
β -D-Gal-(1 $\rightarrow$ 2)-Xyl
 β -D-Gal-(1 $\rightarrow$ 3)-Xyl
 β -D-Gal-(1 $\rightarrow$ 4)-Xyl

Enzymatic synthesis, isolation, and analysis of novel α - and β -galactosyl-cycloisomalto-octaoses

Kyoko Koizumi ^a, Toshiko Tanimoto ^a, Yoko Kubota ^a, Sumio Kitahata ^b

^a School of Pharmaceutical Sciences, Mukogawa Women's University, 11-68 Koshien Kyuban-cho, Nishinomiya 663, Japan

^b Osaka Municipal Technical Research Institute, 1-6-50 Morinomiya, Jyoto-ku, Osaka, 536, Japan



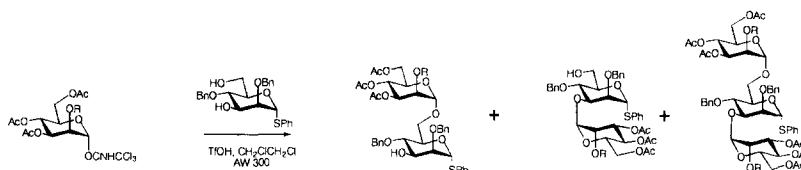
and their β -galactosyl isomers [β -Gal-Clg (I), (II)]

Chemoenzymatic synthesis of the branched oligosaccharides which correspond to the core structure of N-linked sugar chains

Ichiro Matsuo ^a, Megumi Isomura ^a, Tatsuo Miyazaki ^b, Tohru Sakakibara ^b, Katsumi Ajisaka ^a

^a Meiji Institute of Health Science, Meiji Milk Products Co., Ltd., 540 Naruda, Odawara 250, Japan

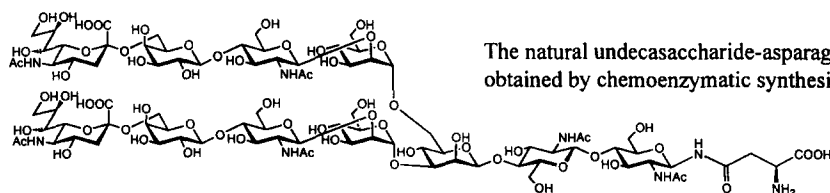
^b Department of Chemistry, Yokohama City University, Seto, Kanazawa-ku, Yokohama 236, Japan



Chemoenzymatic synthesis of a sialylated diantennary N-Glycan kinked to asparagine

Carlo Unverzagt

Institut für Organische Chemie und Biochemie, TU München, Lichtenbergstr. 4, D-85748 Garching, Germany



The natural undecasaccharide-asparagine **1** was obtained by chemoenzymatic synthesis

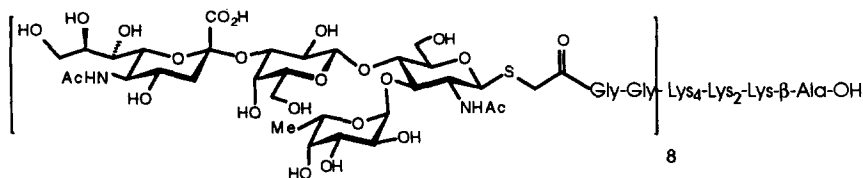
1

Chemoenzymatic synthesis of dendritic sialy Lewis^x

Monica M. Palcic^a, Hong Li^a, Diana Zanini^b, Resham S. Bhella^a, René Roy^b

^a Department of Chemistry, University of Alberta, Edmonton, AB Canada T6G 2G2

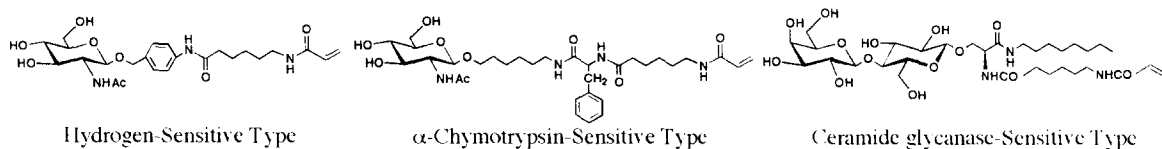
^b Department of Chemistry, University of Ottawa, Ottawa, ON Canada K1N 6N5



High performance polymer supports for enzyme-assisted synthesis of glucoconjugates

Kuriko Yamada, Eriko Fujita, Shin-Ichiro Nishimura

Division of Biological Sciences, Graduate School of Science, Hokkaido University, Nishi-8, Kita-10, Kitaku, Sapporo 060, Japan



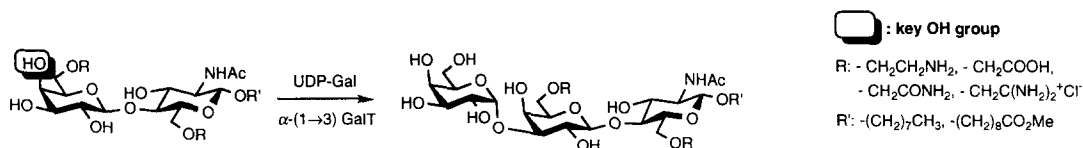
Acceptor hydroxyl group mapping for calf thymus

α -(1 $\rightarrow$ 3)-galactosyltransferase and enzymatic synthesis of α -D-Galp-(1 $\rightarrow$ 3)- β -D-Galp-(1 $\rightarrow$ 4)- β -D-GlcNAc analogs

Keiko Sujino^{a,b}, Carles Malet^a, Ole Hindsgaul^a, Monica M. Palcic^a

^a Department of Chemistry, University of Alberta, Edmonton, Alberta T6G 2G2 Canada

^b The Noguchi Institute, 1-8-1, Kaga, Itabashi-ku, Tokyo 173 Japan



Enzymatic synthesis of site-specifically α 1-3-fucosylated polylactosamines containing either a sialy Lewis X, a VIM-2, or a sialylated and internally difucosylated sequence

Jarkko Rääbinä^{a,b}, Jari Natunen^a, Ritva Niemelä^a, Heidi Salminen^a, Kristiina Ilves^a, Olli Aitio^{a,c}, Hannu Maaheimo^a, Jari Helin^a, Ossi Renkonen^a

^a Inst. of Biotechnology and Dept. of Biosciences (Division of Biochemistry), University of Helsinki, Finland

^b Haartman Inst., Dept. of Bacteriology and Immunology, University of Helsinki, Finland

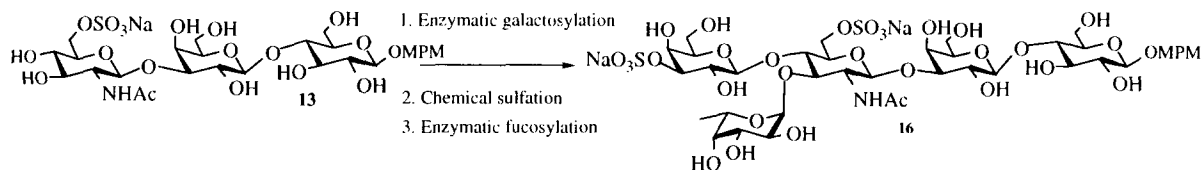
^c Dept. of Pharmacy, Division of Pharmaceutical Chemistry, University of Helsinki, Finland

Enzymatic synthesis of hexasaccharides Neu5Ac(α 2-3)Gal(β 1-4)GlcNAc(β 1-3)Gal(β 1-4)[Fuc(α 1-3)]GlcNAc and Neu5Ac(α 2-3)Gal(β 1-4)[Fuc(α 1-3)]GlcNAc(β 1-3)Gal(β 1-4)GlcNAc, and nonasaccharide Neu5Ac(ga2-3)Gal(β 1-4)GlcNAc(β 1-3)Gal(β 1-4)[Fuc(α 1-3)]GlcNAc(β 1-3)Gal(β 1-4)[Fuc(α 1-3)]GlcNAc is described.

Chemo-enzymatic synthesis of a 3^{IV},6^{III} disulfated Lewis^x pentasaccharide, A candidate ligand for human L-selectin

André Lubineau, Claudine Augé, Nicole Le Goff, Christine Le Narvor

Institut de Chimie Moléculaire d'Orsay, U.R.A. C.N.R.S. 462, Université de Paris-Sud, Bt 420, F-91405, Orsay, France



Fucosyltransferase-catalyzed formation of L-galactosylated Lewis structures

Katja Stangier ^{a,b}, Monica M. Palcic ^a, David R. Bundle ^a, Ole Hindsgaul ^a, Joachim Thiem ^b

^a Department of Chemistry, University of Alberta, Edmonton, Alberta T6G 2G2, Canada

^b Institut für Organische Chemie, Universität Hamburg, D-20146 Hamburg, Germany

α 1-3/4 FucT could be employed for L-galactosylation of disaccharide acceptor substrates with (modified) GDP-L-Gal to give (modified) L-galactosylated Le^a and Le^x trisaccharide derivatives.

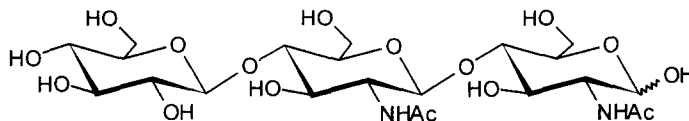
β -Glucosylation of chitooligomers by galactosyltransferase

Vladimír Křen ^a, Jana Dvořáková ^a, Ulrike Gamber ^b, Petr Sedmera ^a, Vladimír Havlíček ^a, Joachim Thiem ^b, Karel Bezouška ^c

^a Institute of Microbiology, Czech. Acad. Sci., Laboratory of Biotransformation, CZ 142 20 Prague 4, Czech Republic

^b Institute of Organic Chemistry, University of Hamburg, D-20146, Germany

^c Faculty of Science, Charles University Prague, Dept. of Biochemistry, CZ 128 40 Prague 2, Czech Republic



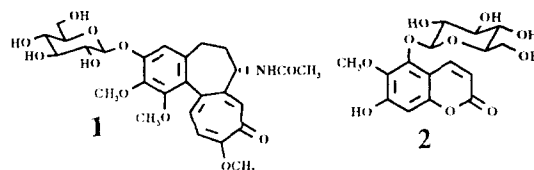
Effects of organic cosolvents on the stability and activity of the β -1,4-galactosyltransferase from bovine colostrum

Sergio Riva ^a, Barbara Sennino ^a, Francesca Zambianchi ^a, Bruno Danieli ^b, Luigi Panza ^b

Istituto di Chimica degli Ormoni, C.N.R., Via Mario Bianco 9, I 20131 Milano, Italy

Dipartimento di Chimica Organica ed Industriale, Università di Milano, Via Venezian 21, I 20131 Milano, Italy

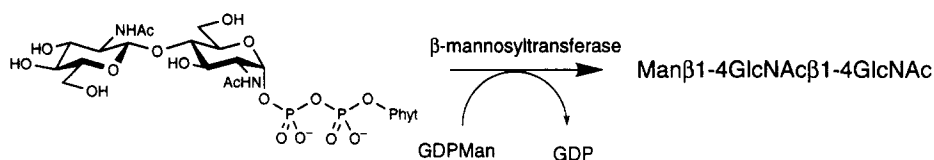
The influence of different organic cosolvents on the stability and activity of the β -1,4-galactosyltransferase from bovine colostrum and of its ancillary enzyme UDP-galactose-4'-epimerase has been investigated using the alkaloid glucoside colchicoside (**1**) and the coumarinic glucoside fraxin (**2**) as model substrates.



The chemoenzymatic synthesis of the core trisaccharide of N-linked oligosaccharides using a recombinant β -mannosyltransferase

Gregory M. Watt, Leigh Revers, Matthew C. Webberley, Iain B.H. Wilson, Sabine L. Flitsch

The Edinburgh Centre for Protein Technology, Department of Chemistry, King's Buildings, The University of Edinburgh, West Mains Road, Edinburgh EH9 3JJ, UK



Structural characterization of the maltose acceptor products synthesized by *Leuconostoc mesenteroides* NRRL B-1299 dextranucrase

Marguerite Dols ^a, Magali Remaud-Simeon ^a, René-Marc Willemot ^a, Michel R. Vignon ^b, Pierre Monsan ^a

^a Centre de Bioingénierie Gilbert Durand, UMR. CNRS 5504 - LA. INRA, INSA Complexe Scientifique de Rangueil, F-31077 Toulouse, France

^b CERMAV-CNRS, F-38041 Grenoble, France

The structure of three different families of oligosaccharides synthesized during acceptor reaction with maltose and sucrose by *L. mesenteroides* NRRL B-1299 dextranucrase was solved. A cascade scheme for the biosynthetic pathway is proposed.

Transfer reactions catalyzed by cyclodextrin glucosyltransferase using 4-thiomaltosyl and C-maltosyl fluorides as artificial donors

Laurent Bornaghi ^a, Jean-Pierre Utile ^a, El Djouhar Rekaï ^b, Jean-Maurice Mallet ^b, Pierre Sinaÿ ^b, Hugues Driguez ^a

^a Centre de Recherches sur les Macromolécules Végétales, CNRS, B.P. 53, 38041 Grenoble cedex 9, France

^b Ecole Normal Supérieure, Département de Chimie, 24 rue Lhomond, 75231 Paris cedex 05, France

